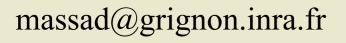
Plant Atmosphere ammonia exchange: a modeling frame to include plant metabolism



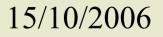
MASSAD Raia Silvia LOUBET B., TUZET A. and CELLIER P.

INRA - Environment and Arable Crops Research Unit



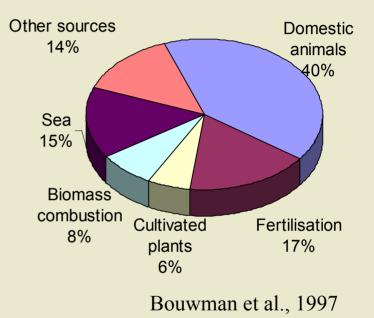






Why study vegetationatmosphere ammonia exchange

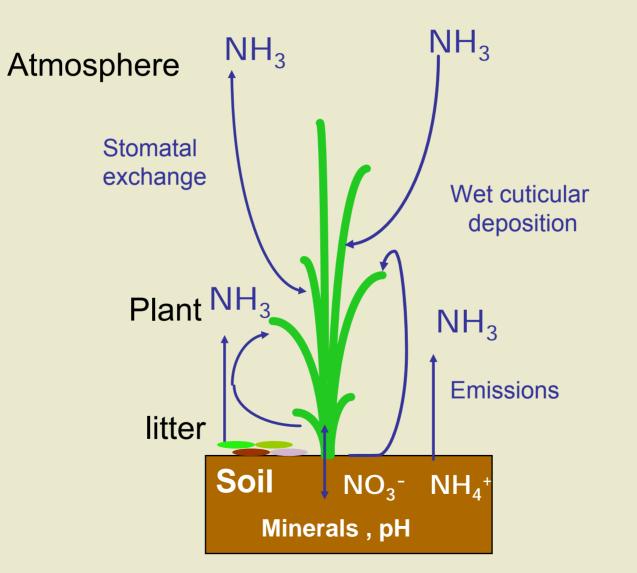
- Quantitatively emission and deposition fluxes are poorly defined
- Important gaps in the mechanistic understanding of the exchange process



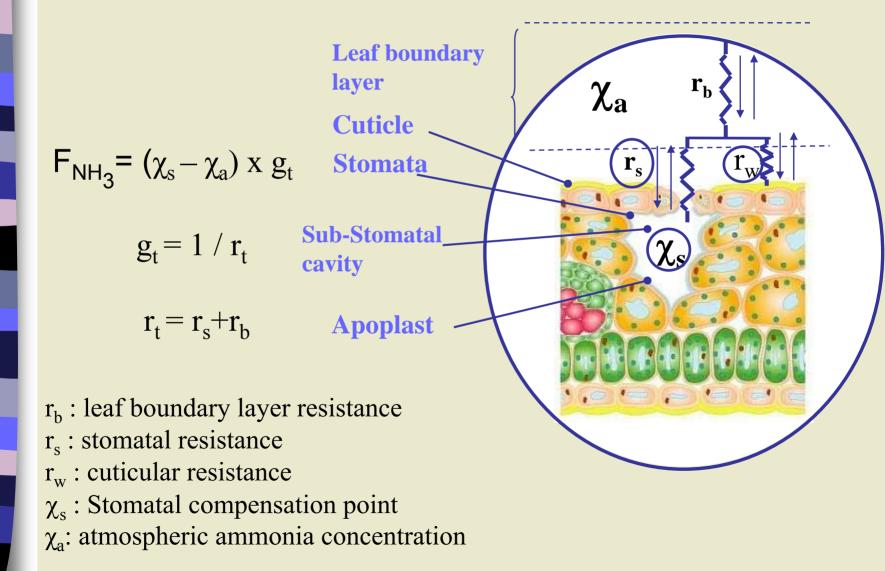
 Livestock sources dominate point source emissions, evidence suggests that exchange with vegetation plays a major role in regulating both air concentrations and the extent of long-range transport (Langford and Fehsenfeld 1992; Sutton et al. 1994)



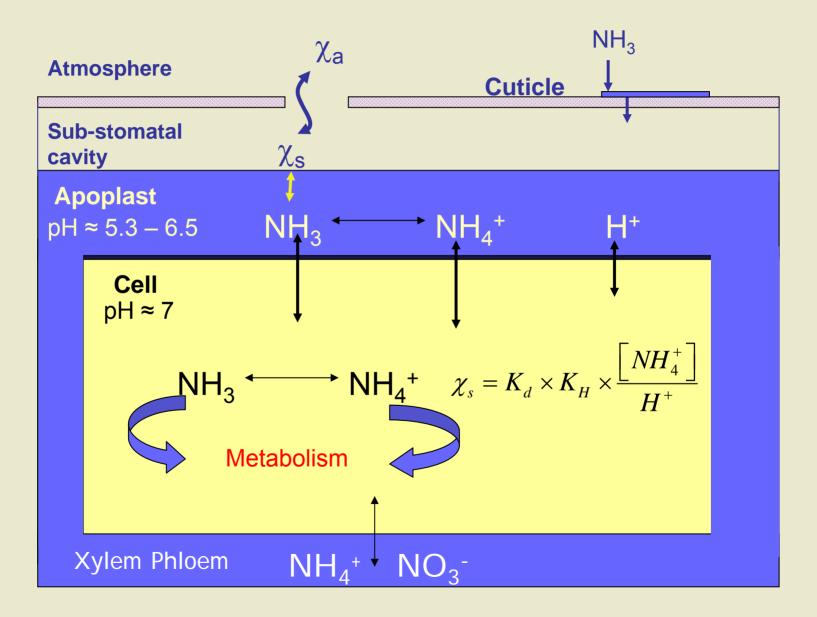
Vegetation-atmosphere NH₃ exchange



Leaf-atmosphere NH₃ exchange



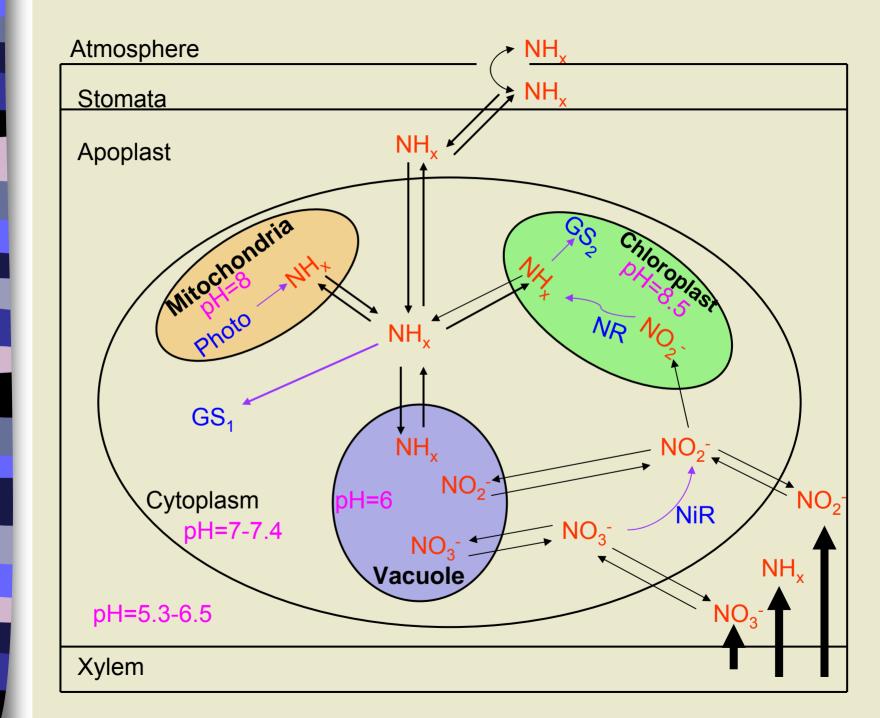
Cell-atmosphere NH₃ exchange

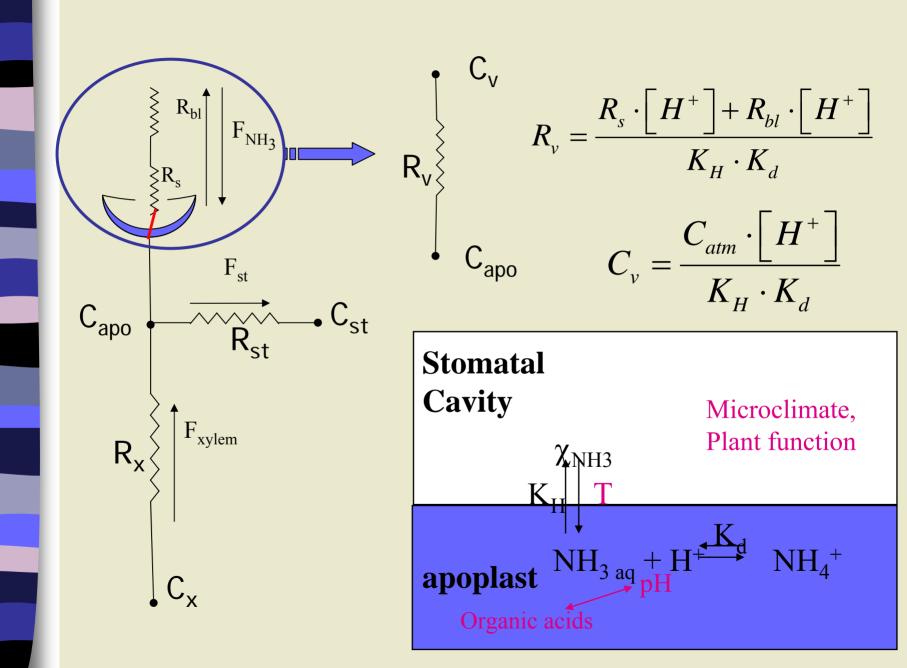


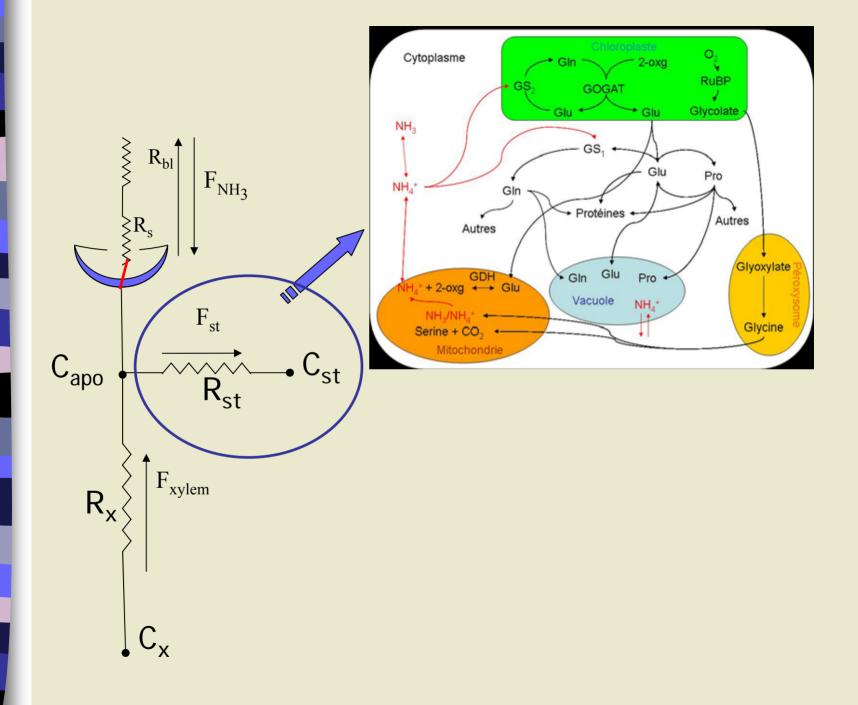
Cell metabolism & NH₃ exchange

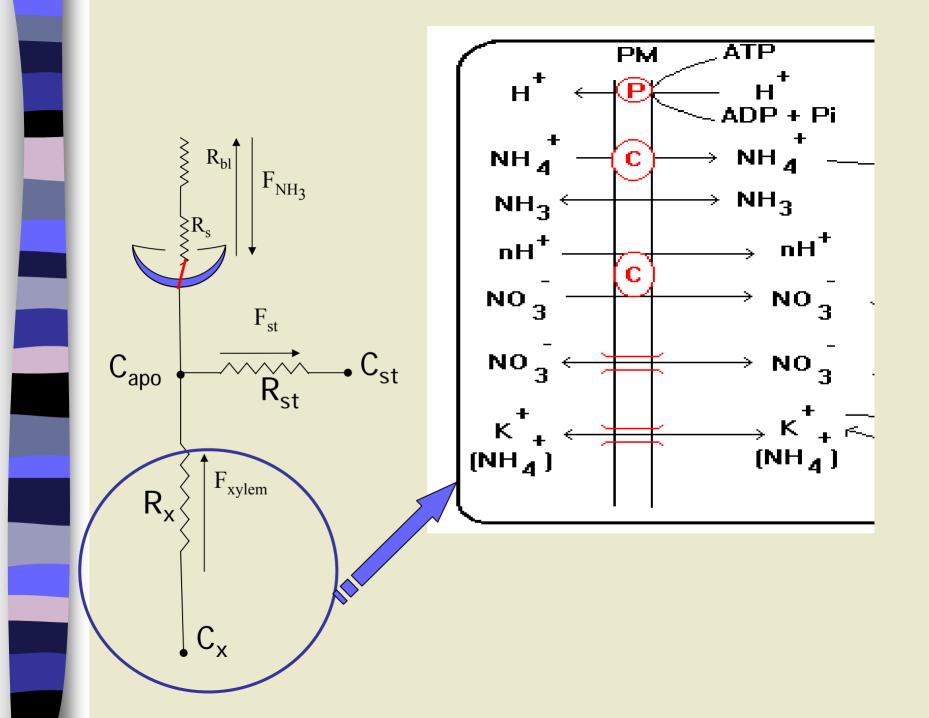
- [NH₄⁺]_{apoplast} depends on:
 - > the balance of import via the xylem,
 - > the absorption by cells,
 - \geq and the export by the phloem.
- Assimilation occurs mainly via the GS/GOGAT cycle
 - Production occurs via:
 - > nitrate reduction,
 - > photorespiration,
 - > ureic catabolism,
 - lignin synthesis,

> decomposition of glutamine and asparagine,









Objectives

- Identify limiting processes in determining [NH₄⁺]_{apo}
- Maintain a mechanistic approach
- Account for N-nutrition, environmental conditions, plant metabolism

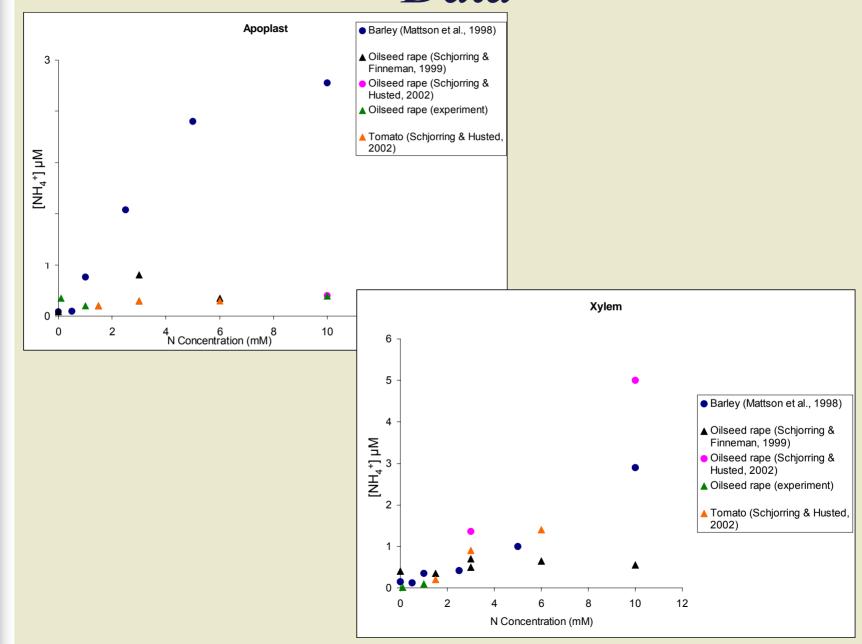
Procedure

- Step 1: Collect data and do some experimentation
- Step 2: Conceptually validate each hypothesis
- Step 3: Estimate resistance values by combining data and equations
- Step 4: Use estimated resistances to calculate apoplastic concentrations and compare

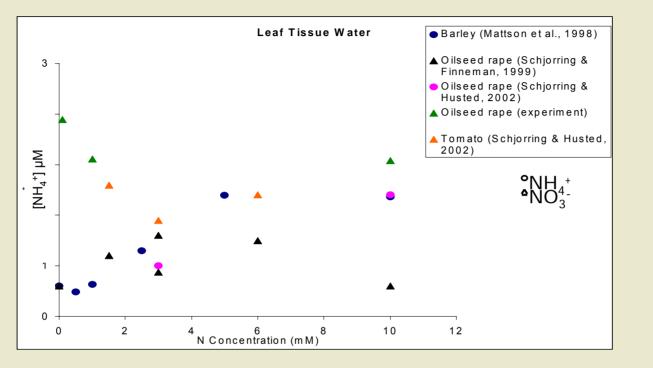
Experiment

- Oil seed rape in phytotron with controlled conditions and grown hydroponically with different N treatments (0,1 mM NO₃⁻, 1 mM NO₃⁻, 10 mM NO₃⁻ and 5 mM NH₄⁺)
- Extraction and measurement of ammonia concentrations and pH for:
 - apoplast by infiltration/centrifugation technique (Husted & Schjørring, 1995),
 - xylem
 - Leaf tissue water
- Photosynthesis measurements
- Total Carbon and Nitrogen contents

Data



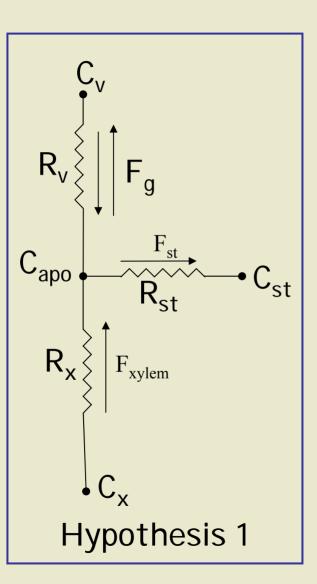
Data



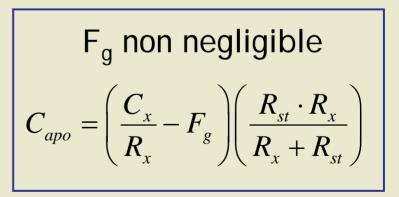
 $F_x = C_x \times ET$ $F_{st} = A \times N/C$ $F_{NH_3} = (C_s - C_a) \times g_s$

F _x (μmol m ⁻² s ⁻¹)	F _{st} (μmol m ⁻² s ⁻¹)	F _g (μmol m ⁻² s ⁻¹)
0.07	1.43	0.04
0.06	1.70	-0.02
0.14	2.60	0.01

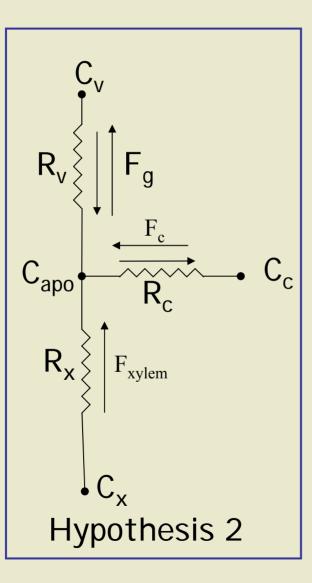
Model equations



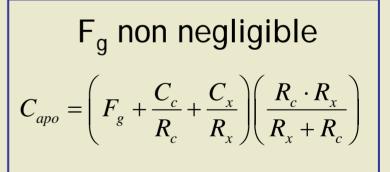
$$F_{g} \text{ negligible}$$
$$C_{apo} = C_{x} \left(\frac{R_{st}}{R_{x} + R_{st}} \right)$$



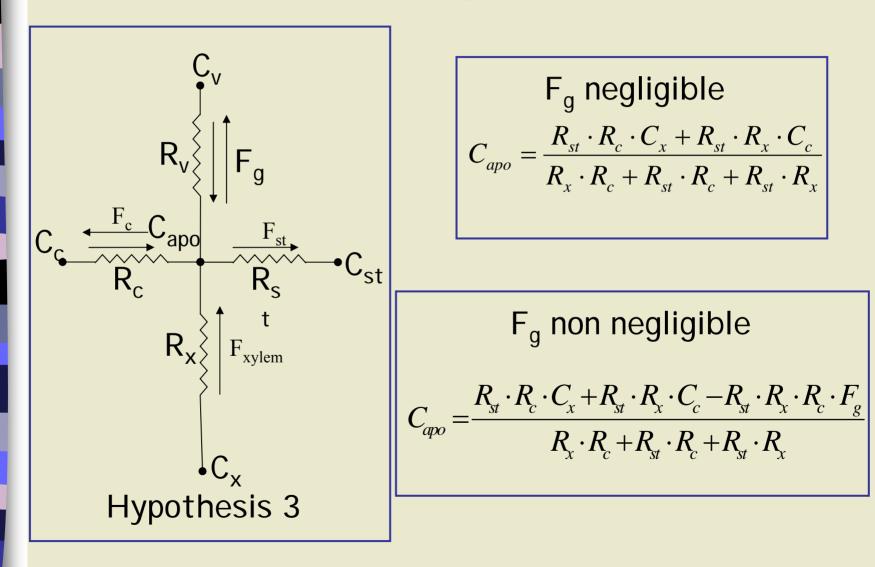
Model equations



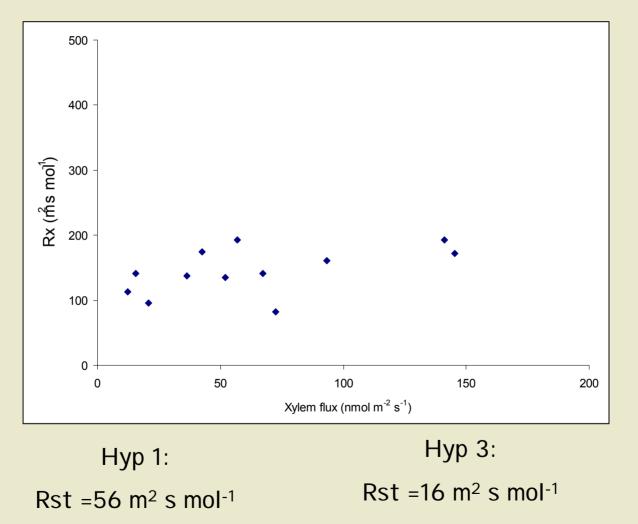
$$F_{g} \text{ negligible}$$
$$C_{apo} = \left(\frac{C_{x}}{R_{x}} + \frac{C_{c}}{R_{c}}\right) \left(\frac{R_{c} \cdot R_{x}}{R_{x} + R_{c}}\right)$$



Model equations

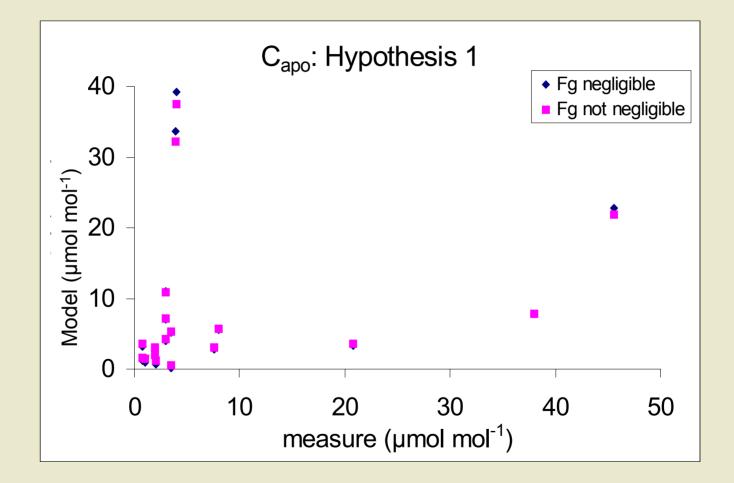


Estimation of Resistances

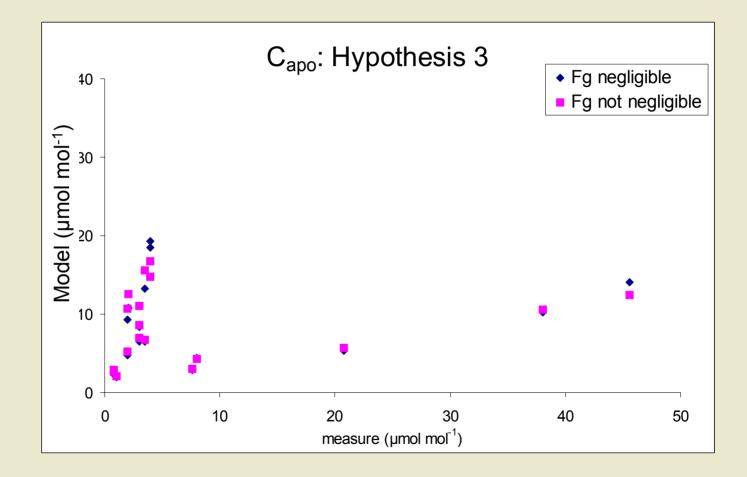


 $Rc = 26 m^2 s mol^{-1}$

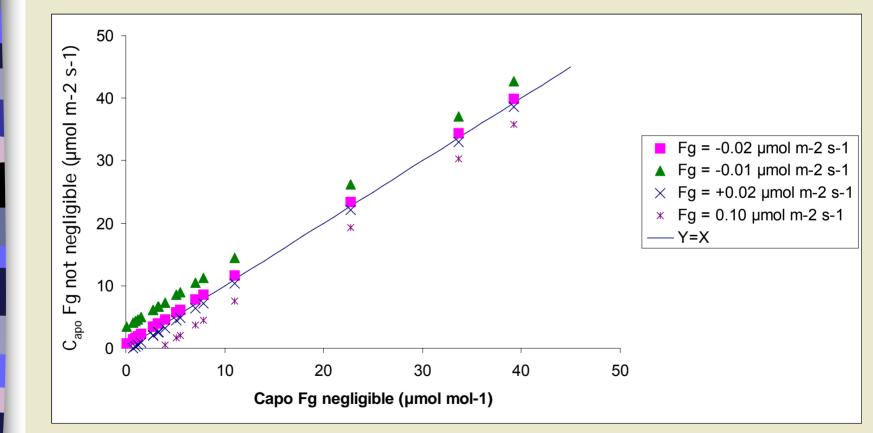
Hypothesis 1: Results



Hypothesis 3: Results



Effect of F_g on C_{apo}





Conclusions & Perspectives

- Possible explanation for difference between Flux measurement and extraction techniques
- Conceptually acceptable but doesn't account for some biological realities
- Have more adapted set of data
 - Flux measurements (Isotopic tracers)
 - Dynamic measurements
- Relate to Nitrate nutrition
- Relate resistances to biological functioning
- Integrate senescence through possibility of back flow from C_{st}
- Link to SVAT models